

Blastocyst development in assisted reproductive technologies: a narrative review evaluating its role as a surrogate marker for pregnancy outcomes and live birth success

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There is an urgent global need to improve in vitro fertilization success rates and expand access to services. Specific to the in vitro fertilization laboratory, challenges such as standardization and a shortage of trained embryologists hinder quality and limit service availability. Current standards for product approval rely on demonstrating comparable pregnancy rates, requiring extensive patient involvement and time-consuming trials, which may be further hindered by patient reluctance to participate in clinical trials. Efficient assessment of new protocols and devices for assessing human assisted reproductive technology requires considering intermediate endpoints and markers to complement conventional endpoints. This review explores blastocyst development as a potential surrogate marker for pregnancy. It examines the correlation between blastocyst development and implantation potential, evaluates how culture conditions and other factors affect outcomes, and discusses the evidence supporting an absence of adverse effects of embryo culture on perinatal and offspring health. The conclusion strongly suggests that blastocyst development could serve as a valuable surrogate for establishing equivalency of pregnancy and live births in new assisted reproductive technology protocols. This review underscores the need for a surrogate marker of quality and presents evidence supporting the utility of blastocyst use rate as a sufficient indicator. (F S Rev® 2025;6:100094. ©2025 by American Society for Reproductive Medicine.)

Key Words: Assisted reproductive technology, innovation, blastocyst development

ESSENTIAL POINTS

- Blastocyst development rate is sensitive to suboptimal conditions, occurring after embryonic genome activation and correlating strongly with both aneuploidy and implantation potential.
- Blastocyst development rates show consistent statistical inference with pregnancy outcomes—specifically, reduced blastocyst rates are correlated with reduced pregnancy rates, whereas equivalent or increased rates are associated with equivalent pregnancy rates.
- Using blastocyst development as a surrogate endpoint for regulatory approval of new devices and protocols could accelerate innovation in assisted reproduction while maintaining safety standards, particularly for automation technologies aimed at improving laboratory standardization and efficiency.

Fertility clinics face significant challenges in delivering high-quality care due to increasing demand coupled with a shortage of skilled embryological professionals. Factors such as delayed childbearing and

increasing infertility rates contribute to the growing demand for fertility services, straining resources and leading to longer wait times for treatment. Meanwhile, high costs associated with in vitro fertilization (IVF) procedures

create disparities in access, which is exacerbated by a shortage of specialized healthcare professionals. Technological innovation in the IVF laboratory provides a strategy for addressing these challenges, by developing tools

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to standardize and optimize protocols (1–4), thereby improving access (5), and extending the fertility care workforce (2, 5). Most products used in the IVF laboratory were established before 2000, and newer introductions often reference the Food and Drug Administration (FDA)'s Final Rule, 63 FR48428 for 510K approval. Notably, protocols such as intracytoplasmic sperm injection (ICSI) (6) and embryo cryopreservation (7) have evolved over time with minimal FDA oversight. In contrast, novel device development necessitates *de novo* applications to the FDA, with safety and efficacy traditionally evaluated on the basis of live birth rates, a metric requiring substantial sample sizes and protracted timelines for recruitment and evaluation. Given the burdens—financial and emotional—borne by patients with infertility, participation in clinical trials is often constrained, impeding innovation in new product development and the opportunity for enhancement of fertility treatment.

Innovations in assisted reproductive technology (ART) can be categorized into several distinct groups, each with varying potential to benefit from blastocyst development as a surrogate marker. Culture system technologies include the following: culture media formulations (conventional single-step/sequential media, protein supplements, and media with molecules such as antioxidants); physical culture parameters (oxygen concentration, temperature, pH, and humidity); and culture vessels and accessories (dishes, oil overlays, and incubation systems). Automation technologies represent a rapidly developing sector encompassing oocyte handling systems (retrieval and denudation), sperm processing and ICSI automation, embryo culture systems (time-lapse imaging), and cryopreservation automation (vitrification devices). Diagnostic and selection tools include noninvasive embryo assessment (morphokinetics, metabolomics, and spent media analysis), genetic testing methodologies (preimplantation genetic testing for aneuploidy and emerging noninvasive genetic testing), and artificial intelligence and machine learning platforms for embryo selection. Automation technologies are particularly relevant for the use of blastocyst development as a surrogate marker because these innovations primarily aim to replicate current manual processes with equivalent outcomes rather than necessarily improving pregnancy rates. Establishing equivalence rather than superiority is the appropriate standard for these technologies.

To expedite new procedural and device development for fertility treatments, the adoption of an intermediate endpoint capable of accurately assessing risk and efficacy holds promise in shortening approval timelines, thereby accelerating generalized introduction of new fertility treatments to clinical care. Surrogate endpoints, well-established alternatives, streamline trial efficiency while furnishing requisite safety and efficacy data for FDA scrutiny (8, 9). Over the past 3 decades, numerous biomarkers or intermediate outcomes have gained acceptance as surrogate endpoints in regulatory approvals for pharmaceuticals and medical devices (10, 11). Notably, research in IVF routinely uses surrogate endpoints for live birth, such as blastocyst development and clinical/on-going pregnancy rates (12–17). In this review, we consider

whether blastocyst development holds promise as a suitable endpoint for clinical trials.

Piantadosi (18) outlines key attributes of a valuable surrogate endpoint: ease of measurement without invasive procedures; relevance to the causal pathway for the preferred endpoint; consistent statistical inference; and responsiveness to treatment effects. Mere correlation between a surrogate and clinical outcome falls short; instead, the intervention's impact on the surrogate should reliably predict its effect on the clinical outcome. Considering the critical stages of human embryo development, this review concentrates on blastocyst development as a pivotal stage and probes its potential as a surrogate endpoint for pregnancy rates, applying Piantadosi's (18) criteria for evaluation.

This narrative review examines the evidence supporting blastocyst development as a surrogate endpoint for pregnancy and live birth in ART using literature identified through searches of PubMed/MEDLINE using combinations of terms related to assisted reproduction (“in vitro fertilization” and “IVF”), blastocyst development (“blastocyst” and “embryo culture”), clinical outcomes (“pregnancy rate” and “live birth”), and surrogate endpoints (“surrogate marker” and “validation”). Additional relevant studies were identified through reference list screening and citation tracking of key articles. The review incorporates 3 decades of research and clinical experience in human ART, critically evaluating the strength of evidence supporting blastocyst development as a surrogate marker. Although animal studies are referenced where relevant, particularly regarding developmental biology and safety assessment, the primary focus is on human clinical outcomes. Evidence is assessed for quality and relevance, with careful distinction made between established findings and expert interpretation. This approach aims to provide a comprehensive evaluation of whether blastocyst development could serve as a reliable surrogate endpoint while maintaining the high safety standards essential in reproductive medicine.

BLASTOCYST DEVELOPMENT: AN EASILY MEASURED SURROGATE THAT IS PART OF THE CAUSAL PATHWAY OF IMPLANTATION AND CAN BE ASSESSED NONINVASIVELY WITHOUT IMPAIRING FUTURE POTENTIAL FOR SUCCESS

Clinical IVF laboratories are unique in medicine in that they provide the safe, *ex vivo*, passage of gametes and embryos—the foundations of generational health. Although the original protocols and products used in clinical IVF were nearly exclusively developed in animal models with few safety studies in humans (19), today's modern IVF laboratory evaluates different protocols and products for quality improvement initiatives as part of a broader quality management system (QMS) (20–23). Markers of efficacy vary from those specific to a developmental stage (e.g., oocyte maturation rate and fertilization rate) to markers that reflect the entire ecosystem of the IVF laboratory (e.g., blastocyst development). The goal of a QMS approach to the

assessment and implementation of change in the laboratory is to establish equivalence before the introduction into routine practice, without the need to wait for live birth rates (24).

Laboratory automation, whereby an automated method replaces a manual one, also requires proof of equivalence. For instance, some intermediate markers such as accuracy of automated semen analysis (25), recovery rate of microfluidic sperm isolation (26), or survival after embryo automated vitrification (27) are essential to confirm safety and efficacy before implementation. However, these intermediate measures do not assess the impact of the change on the entire process and, ultimately, on success of fertility treatment.

The rate at which fertilized oocytes develop into usable blastocysts, or the blastocyst use rate (BUR), is the most comprehensive measure of an embryo culture ecosystem. Unlike other intermediate measures, such as fertilization rate and cleavage stage development, usable blastocyst rate depends on embryonic genome activation (28) and is responsive to aneuploidy (i.e., blastocyst rate is reduced for aneuploid embryos (29)). Although there is a direct relationship between blastocyst grade (30), speed of development (31), and implantation rate for a single embryo transfer, the development of a cohort of fertilized oocytes from each patient provides a robust measure of overall laboratory performance.

To be a trusted surrogate marker for equivalency of a treatment, blastocyst development rate requires a standardized definition. The Alpha/European Society of Human Reproduction and Embryology consensus on embryo grading provides 2: number of good-quality blastocysts on day 5 and total blastocysts on day 5 (32, 33). These key performance indicators require further development for 2 reasons: they do not incorporate total blastocyst development rate to day 6 and/or 7, and they do not provide a definition for “good quality.” Most publications address the issue of quality by presenting the percentage of top-quality blastocyst (34), typically those with grade A inner cell mass and/or trophectoderm (TE) on the basis of the Gardner grading scheme (35). Similarly, the total number of blastocysts is based on the number used, either transferred or cryopreserved (36). Although blastocyst grading is subjective (37) and definitions of usability vary among clinics (38), good-quality blastocyst formation, usable development rate, and overall BUR are effective when comparing 2 or more methods *within* a laboratory. Although different measures have merit, this review will use the term BUR as a general term for rate of blastocyst development.

Several potential surrogate markers for pregnancy and live birth, which can be obtained easily and noninvasively, are present during human embryo in vitro culture’s developmental timepoints. Most preimplantation developmental stages have been correlated with implantation potential (39–44). However, an effective surrogate should occur after human embryonic genome activation, which occurs during the cleavage stages of development (days 1–3 (45)). Furthermore, as blastocyst culture has become a part of routine clinical practice, evidence suggests that predictive features before embryo compaction reflect an embryo’s ability to develop in vitro and that these early features lose

predictive ability for implantation after blastocyst formation (46, 47).

The speed and quality of blastocyst development have been repeatedly demonstrated to be strongly correlated with the chance of live birth. Blastocysts that develop “on time” or early (48) have significantly higher implantation rates than those that develop on day 6, which are higher than those developing on day 7 (31, 49–53). Coupled with speed, blastocyst morphology is associated with implantation potential, with the TE grade showing the highest correlation with live birth (54–56). Blastocyst size (55, 57, 58) and inner cell mass grade (59, 60) also have associations with live birth. These 3 variables are compiled into 1 score, for example, the Gardner blastocyst grade (61), and, when coupled with developmental speed, provide meaningful prognostic insights into a patient’s chance for a live birth (29). The relationship between blastocyst quality and implantation is robust, leading to development of artificial intelligence-based embryo selection systems (62–70).

The rate of euploid blastocyst development could be considered a better surrogate endpoint than BUR without genetic testing; however, it is invasive and, thus, does not meet one of Piantadosi’s (18) criteria. Preimplantation genetic testing for aneuploidy is an objective diagnostic assessment of an embryo’s developmental potential. It provides insights into blastocyst potential, and thus, euploid blastocyst rates could also as serve as a surrogate endpoint for pregnancy. Furthermore, assessment of embryo genetics in relation to pregnancy outcomes would be beneficial, particularly if such assessments could be obtained noninvasively (71–73).

Many early developmental milestones that are correlated with blastocyst development are also correlated with aneuploidy. For instance, morphokinetic algorithms show good prediction of both aneuploidy and pregnancy (74–76). A strong relationship exists between aneuploidy and blastocyst developmental speed and quality (77). Aneuploidy is higher for slow growing day 6 and 7 blastocysts (49, 78, 79) and for lower-grade blastocysts (80). It is unlikely that earlier developmental stages provide additional prognostic value over blastocyst development alone or euploid blastocyst development.

The number of blastocysts or euploid blastocysts obtained from mature oocytes or zygotes is the euploid BUR. The euploid BUR is likely the best surrogate for pregnancy when testing a proposed or alternate procedure or intervention because it can detect mitotic aneuploidies that occur during embryo culture, aneuploidies that may be the result of an intervention (81). Although promising in theory, evidence for an impact of culture on rates of aneuploidy is currently limited (81, 82). Furthermore, until a noninvasive genetic test is developed, this metric requires an invasive TE biopsy. Although blastocyst quality and speed of development are associated with aneuploidy, implantation potential of euploid blastocysts diminishes the impact of quality and speed of development on implantation rates (78, 83).

In summary, the rate of good-quality blastocyst development per oocyte/zygote is independently the strongest, and perhaps the only, noninvasive predictor of implantation

TABLE 1

Summary of retrospective and prospective studies of associations between chemical and physical parameters and blastocyst rate and pregnancy rate, adapted from the study by Bartolacci et al. (90).

Parameter	Comparison	Reference group	Studies	RCTs with both BR and PR	Studies with statistical inference for equivalence
Temperature	37 vs. <37 °C	37 °C	5 (91–95)	3	3 (93–95)
Oxygen	5% vs. 20%	5%	9 (96–104)	5	5 (100–104)
Oxygen	Biphasic (5–2%) vs. 5%	Biphasic	9 (105–113)	3	3 (111–113)
Humidity	Humid vs. dry	Humid	5 (114–118)	1	1 (114)
Incubator	Time-lapse vs. standard	Time-lapse	7 (119–125)	3	3 (122–124)

Note: BR = blastocyst rate; PR = pregnancy rate; RCT = randomized controlled trial.

Morbeck. Blastocyst rate - a surrogate marker. F S Rev 2025.

within the causal pathway of in vitro embryo development. Only the addition of invasive genetic testing of blastocysts potentially yields a better surrogate.

BLASTOCYST RATE YIELDS THE SAME STATISTICAL INFERENCE AS PREGNANCY RATE AND IS RESPONSIVE TO TREATMENT EFFECTS

Assessing a surrogate marker involves determining whether the statistical analysis of the surrogate provides the same inference about the treatment effect on the true endpoint. Several tests and conditions gauge the degree of similar inference for surrogate endpoints, including the proportion of treatment effect explained (84), consistency of effect across studies (85), and presence of consistency over time for longitudinal surrogates (86). However, the last criterion is nonapplicable in this context because blastocyst rate is not a longitudinal intermediate measure.

A treatment intervention is typically assessed for either equivalence or superiority. For most laboratory process changes or improvements, whether changing to a new culture medium, culture dish, incubator, or automation, establishing equivalence is essential and a requirement of the QMS. In this context, automation—that is, the replacement of a manual method—similarly should establish equivalence and is not required to establish superiority. This is a critical distinction, because the ability to improve pregnancy and live birth rates is constrained by the complexity of the system, which includes variables such as age, the quality and number of gametes, effectiveness of the laboratory, embryo transfer, and uterine receptivity. This concept applies to other areas of medicine, such as the approval of biosimilar drugs, where equivalence, not superiority, is required (87, 88).

When assessing the effect of an intervention on ART outcome, we propose that BUR provides statistical inference for equivalence. Simply put, a treatment leading to similar or better BUR yields similar or better pregnancy rates, whereas a treatment leading to a lower BUR reduces pregnancy rates. This argument applies to cumulative live birth rate from 1 oocyte retrieval and not pregnancy rate per transfer because the surrogate is a measure of the embryo creation cycle. Blastocyst use rate is a critical measure of an intervention's effect

on a patient's entire embryo cohort (89), and thus, an effect on embryo quality that reduces the overall cohort may not be detected by simply following the outcome of the first embryo transfer.

Numerous examples exist that illustrate the relationship between BUR and pregnancy outcomes (Table 1) (90). Chemical and physical features of embryo culture have been assessed in randomized controlled trials, providing examples of the causal relationship between BUR and pregnancy (90). These studies illustrate that the relationship between BUR and pregnancy rate is strong and 1-sided: a reduction in BUR has statistical inference for pregnancy or live birth rate, whereas an equivalent or increased BUR assures an equivalent pregnancy rate (91–125).

Table 1 (90) confirms that numerous laboratory interventions demonstrate a causal relationship for equivalence between blastocyst development and pregnancy rates (90, 126, 127). Although the review (90) included studies on the effects of light (128), features of the oil overlay (115), and pH of culture media (129) and the statistical inference held true, the studies did not include sufficient detail to be included in the summary. Importantly, to our knowledge, there are no examples where an intervention lowers pregnancy rates while not adversely affecting blastocyst development.

Comparing different culture conditions is the most studied laboratory intervention in IVF. Typically, the impact of a change has been assessed using blastocyst development as a primary endpoint and pregnancy outcomes as the secondary endpoints. Culturing in reduced (5%) vs. ambient (20%) oxygen yields more blastocysts and either equivalent or improved pregnancy rates (101, 103, 113, 130, 131). Application of a biphasic oxygen culture, where O₂ is changed from 5% to 2% on day 3, did not change BUR or CPR (111). Culturing embryos in groups vs. individually similarly increases the number and quality of blastocysts while either not affecting or improving pregnancy rates (132, 133). Culturing at a reduced temperature (<37 vs. 37 °C) and in dry conditions (vs. humidified incubation) also impacts outcomes and can lead to fewer blastocysts and either equivalent (134) or reduced pregnancy rates (114).

The composition of culture media varies considerably among the many brands used for clinical IVF (135–137). Comparisons of the effect of culture media on outcomes

present a similar pattern to studies on culture conditions: an impact on blastocyst development with or without an impact on pregnancy rates (138). Few studies provide cumulative outcomes, making the direct link between embryo quality and pregnancy outcomes difficult. Only a single study demonstrated a reduced day 5 blastocyst rate with 1 culture medium that also corresponded to lower pregnancy rates for both the first transfer and cumulative rates (36).

The final feature indicating similar statistical inference to the endpoint is consistency of the effect across studies and populations. The studies presented in Table 1 (90) and in this discussion are from diverse research groups that consistently show that BUR provides statistical inference for equivalence, highlighting blastocyst development's significance in ART.

REVIEWING THE RISKS OF LONG-TERM EFFECTS OF GAMETE AND EMBRYO HANDLING IN VITRO

Although live birth with the absence of neonatal complications is the current standard endpoint for clinical trials in IVF, long-term impacts on health of ART-conceived offspring remain a crucial area of research (139–141). In fact, neonatal outcomes are important surrogate markers for potential long-term health because preterm birth, small for gestational age, and large for gestational age (LGA) are all associated with childhood and adult health (142–145). Offspring conceived through ART exhibit higher rates of preterm birth, small for gestational age, and LGA (146–151).

Assessing the risks associated with embryo culture and laboratory interventions in the context of neonatal outcomes remains an important area of research. Factors that may contribute to adverse outcomes include maternal/paternal age, underlying infertility, ovarian stimulation, laboratory conditions, and the uterine environment after embryo transfer (146). Underlying infertility plays a significant role in observed differences. For instance, birth defects are increased by 30% for ART offspring (152, 153) compared with 20% for offspring from subfertile women not requiring IVF or ICSI (154). Overall, the absolute increase in the risk of adverse outcomes is small, and a direct link between laboratory conditions has not been definitively established. A theoretical monitoring effect, whereby pregnancies and children resulting from ART are closely monitored and seek healthcare more frequently, has also been proposed (155). In general, IVF-conceived children are healthy and develop normally (140, 156).

Despite challenges in identifying causative factors amid underlying infertility, certain laboratory interventions, such as type of culture media (157), extended culture to the blastocyst stage (158), and embryo cryopreservation (151), have been associated with neonatal outcomes (159). Results remain inconclusive, however, with some studies reporting differences in birth weight associated with culture media (160, 161), whereas others find no significant impact (146). The incidence of LGA is higher after blastocyst and frozen embryo transfers (151), which has been attributed to the

lack of a corpus luteum during programmed replacement cycles (162) and the duration of embryo culture (163). The impact of extended culture to the blastocyst stage remains an active area of debate (158, 164–166).

Longitudinal studies examining the impact of changes in clinical practice on birth weight provide evidence that culture media and duration of culture are unlikely to influence neonatal outcomes. In 2 studies from Boston tracking IVF outcomes over a 18–24 year period, birth weights and other neonatal outcomes remained unaffected by significant changes, such as oxygen concentration, culture media type (changing from a simple human tubal fluid solution to a single-step complete medium), and type of incubator (167, 168). A similar study in the United Kingdom found a gradual increase in birth weight over a 25-year period and an association between culture media type and live birth rate; however, no association was found between culture media and birth weight (168, 169). Regarding epigenetic causation, one of the culture medium studies observed deoxyribonucleic acid methylation differences at birth that resolved by the age of 9 years (159), whereas the second study found no difference in methylation status of newborns from the 2 culture media (170).

The theory that laboratory conditions during human ART have long-term consequences remains a question with insufficient high-quality evidence. Two main factors contribute to the hypothesis that conditions during preimplantation embryo development are linked to adverse neonatal and long-term outcomes: first, the epigenome undergoes extensive reprogramming after fertilization and before implantation (171); second, convincing evidence from animal models, such as ruminants (172) and mice (173), demonstrates the adverse impact of culture on epigenetic changes with enduring effects. The critical reprogramming that occurs during this stage of development, combined with evidence of sensitivity in various species, makes it intuitively appealing to suggest that a link to human parallels exist. A reported increase in imprinting disorders in ART-conceived children provides evidence for a direct cause and effect (174, 175). However, the theory remains unproven due to the absence of case reports providing clear evidence of a direct link and the general challenge of registry-based epidemiological studies for disorders with very low incidence (176).

Although clinical ART continuously strives to develop treatments that improve outcomes, instances where culture conditions were suboptimal or even embryotoxic are opportunities to determine whether poor conditions during preimplantation development can have adverse neonatal outcomes. For instance, as of 2014, most IVF clinics still cultured with atmospheric oxygen (177), yet there is no significant effect of the oxidative environment on offspring when culturing at atmospheric oxygen vs. 5% oxygen (178). Similarly, the metabolic stress of culture in a simple salt solution—what would now be considered a suboptimal culture medium (e.g., Fujifilm Irvine's HTF)—has not shown adverse effects on birth weight (157, 167). Pregnancies from cycles where poor air quality reduced available blastocysts have not produced case series or reports on adverse outcomes (179). Additionally, several recalls of embryotoxic mineral oil containing

peroxides have occurred in the past 2 decades (180, 181) and likely more cases where affected lots were used but not recalled, with no reports of long-term adverse outcomes.

The absence of severe impacts from adverse culture conditions aligns with the all-or-none hypothesis of embryo toxicity, suggesting either that the preimplantation embryo succumbs to poor conditions and does not result in a live birth or that the preimplantation embryo survives without long-term effects on offspring (182). This hypothesis is based on the concept that teratogenicity occurs later in development, during organogenesis (183). Although reassuring, further investigation is needed to fully understand the implications of this hypothesis.

In summary, although suboptimal culture conditions influence preimplantation embryo viability, evidence suggests that laboratory factors do not significantly impact the health of offspring conceived through ART. Continued research will provide a more complete understanding of possible long-term effects and direct mitigation of any potential risks associated with ART procedures.

SURROGATE ENDPOINTS FOR DEVELOPMENT OF DEVICES AND BIOLOGICS

Innovation stands as a pivotal force in enhancing the success rates of IVF procedures (184) and streamlining laboratory operations for heightened efficiency and more consistent quality. Although culture media have seen minimal evolution in recent decades (136, 137, 185), potential improvements, such as the incorporation of growth factors (186, 187), could address existing gaps. Similarly, despite the introduction of time-lapse incubators (188), many ART laboratory procedures remain manual and lack standardization.

Various innovations drive the need for distinct and appropriate endpoints for safety assessment. Notably, interventions targeting gene transcription, such as the addition of growth factors or cytokines (189), demand rigorous testing due to potential epigenetic implications. For instance, the inclusion of serum in preimplantation bovine embryo culture has led to well-documented cases of large offspring syndrome (190), emphasizing the need for stringent evaluation of potent transcription-altering factors in culture media. Although some advancements, such as the incorporation of granulocyte-macrophage colony-stimulating factor, have undergone thorough randomized controlled trials with associated live birth rates (187), others, such as antioxidants, have been introduced on the basis of clinical trials using ongoing pregnancy as the endpoint (191).

In addition to optimization of culture conditions, automation of laboratory procedures holds promise for enhancing efficiency and standardization, potentially assessed using surrogate endpoints such as blastocyst development. The lengthy and hands-on nature of embryologist training limits the workforce available to meet increasing demand. Automation of critical laboratory steps could simultaneously improve reproducibility while significantly reducing training time (and possibly processing time) and facilitating standardization and improved quality of care. Several automation initiatives are underway, encompassing procedures such as oocyte

retrievals (192), cumulus cell removal (193, 194), ICSI (195, 196), embryo culture, and oocyte and embryo cryopreservation (27, 197).

Blastocyst development emerges as a feasible surrogate endpoint for automation due to several factors. The variability in laboratory procedures underscores the adaptability in gamete and embryo manipulation techniques. Automation aims to replicate human actions while maintaining optimal conditions, using components already prevalent in the laboratory setting. Many of the components are already in use in the laboratory, and those that are not can be tested with well-validated embryo toxicity assays (198).

An example of the inherent heterogeneity in the IVF laboratory and the utility of blastocyst development as a surrogate endpoint is the relatively recent evolution and broad application of oocyte freezing. Early success was achieved with slow freezing protocols (199). Since then, vitrification has become standard for both oocytes and embryos (7). During the past 30 years, countless variations of cryoprotectants (200, 201), temperature (202), exposure times (203–205), and devices (202, 206, 207) have been developed and used, most using oocyte survival, fertilization, and embryo development as surrogate endpoints. A recent study even demonstrated that only 2 minutes is needed for equilibration before vitrification (208), a major change from the long-held belief that at least 10 minutes is required for complete equilibration of vitrification solutions (209). The improvement in efficiency for the laboratory is driving a rapid change in clinical practice, and commercially available products that incorporate these changes are now available, all on the basis of predicate applications.

Establishing the safety of materials is a necessary first step, followed by demonstrating that automated processes yield comparable blastocyst development rates, quality, and speed as manual techniques. Given the strong correlation between blastocyst development and pregnancy outcomes, efficacy confirmation using this surrogate endpoint paves the way for broader approval and adoption of automated processes in ART laboratories. Continuous monitoring of pregnancy and birth outcomes remains essential to ensure ongoing safety and efficacy.

REASONS FOR CAUTION WHEN CONSIDERING BLASTOCYST RATE AS A SURROGATE FOR PREGNANCY

Although blastocyst development demonstrates promising correlations with pregnancy outcomes, several factors warrant consideration when evaluating its suitability as a surrogate endpoint for clinical trials and regulatory approval. The relationship between blastocyst formation and successful pregnancy is influenced by multiple factors beyond the developmental competence captured by blastocyst rate alone. Euploid blastocyst implantation rates typically range from 50% to 70% (210, 211), indicating a significant gap between blastocyst formation and pregnancy success. Factors that contribute to this gap include embryonic factors beyond standard morphological assessment (210), maternal factors such as endometrial receptivity (212), paternal contributions to

embryo competence (213, 214), and various clinical and laboratory variables (210). These additional determinants of pregnancy success highlight the complex nature of implantation that may not be fully captured by blastocyst development metrics.

The methodology of studies evaluating extended culture also affects outcome interpretation. Reports analyzing results on a per-transfer basis may yield different conclusions than intention-to-treat analyses that include all cycle starts (215). Cycle cancellations due to poor embryo development or lack of blastocyst formation are important outcomes that should be considered when evaluating the overall impact of laboratory interventions. Regulatory frameworks should account for how study design influences the apparent predictive value of blastocyst development and ensure that selection bias does not lead to overestimation of clinical efficacy.

Laboratory context also plays a significant role in blastocyst development outcomes. Studies have shown that blastocyst formation rates vary across laboratory settings despite similar patient demographics (81, 216). These variations arise from differences in culture systems, incubation conditions, and technical expertise. When evaluating blastocyst rate as a surrogate endpoint, the transferability of findings across different laboratory environments merits attention to ensure that regulatory decisions on the basis of blastocyst outcomes will translate reliably across diverse clinical settings.

Despite these considerations, blastocyst development remains a valuable biomarker of embryo competence with demonstrated correlation to pregnancy outcomes. When considering its role as a surrogate endpoint for regulatory purposes, these additional factors provide important context for determining the appropriate framework. Although initial regulatory decisions may reasonably rely on equivalent blastocyst quality and numbers as an endpoint, postapproval surveillance and/or additional postapproval studies may be an appropriate means to expeditiously approve new technologies while more carefully confirming safety and efficacy with larger data that can be obtained in a typical phase II/III study. This balanced approach may leverage blastocyst rate as an initial indicator of efficiency and safety for laboratory innovations, particularly those aimed at standardization and automation, while maintaining appropriate monitoring of clinical pregnancy and live birth outcomes to confirm that laboratory improvements translate to meaningful benefits for patients.

SUMMARY

Innovation in ART holds tremendous potential for enhancing overall success rates, streamlining laboratory operations and improving access to treatment. From advancements in culture media to the automation of laboratory procedures, these innovations aim to address existing gaps in efficiency and quality. However, ensuring the safety and efficacy of these innovations requires meticulous evaluation, leveraging surrogate endpoints such as blastocyst development. Blastocyst development is a promising surrogate endpoint for assessing both the safety and efficacy of automated processes in ART laboratories. As ART continues to evolve, ongoing monitoring

and evaluation remain crucial to ensure the safety and effectiveness of these innovations in improving fertility treatments.

CRedit Authorship Contribution Statement

Dean E. Morbeck: Writing – review & editing, Writing – original draft, Formal analysis, Data curation, Conceptualization.
Michael P. Diamond: Writing – review & editing, Writing – original draft, Formal analysis.

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During the preparation of this work the authors used Anthropic's Claude to provide critical editing of grammar and readability. After using this tool/service, the authors reviewed and edited the content as needed and take full responsibility for the content of the publication.

REFERENCES

1. Ueno S, Berntsen J, Ito M, Uchiyama K, Okimura T, Yabuuchi A, et al. Pregnancy prediction performance of an annotation-free embryo scoring system on the basis of deep learning after single vitrified-warmed blastocyst transfer: a single-center large cohort retrospective study. *Fertil Steril* 2021;116:1172–80.
2. Hariton E, Pavlovic Z, Fanton M, Jiang VS. Applications of artificial intelligence in ovarian stimulation: a tool for improving efficiency and outcomes. *Fertil Steril* 2023;120:8–16.
3. Fanton M, Nutting V, Solano F, Maeder-York P, Hariton E, Barash O, et al. An interpretable machine learning model for predicting the optimal day of trigger during ovarian stimulation. *Fertil Steril* 2022;118:101–8.
4. Fanton M, Nutting V, Rothman A, Maeder-York P, Hariton E, Barash O, et al. An interpretable machine learning model for individualized gonadotropin starting dose selection during ovarian stimulation. *Reprod Biomed Online* 2022;45:1152–9.
5. Hariton E, Alvero R, Hill MJ, Mersereau JE, Perman S, Sable D, et al. Meeting the demand for fertility services: the present and future of reproductive endocrinology and infertility in the United States. *Fertil Steril* 2023;120:755–66.
6. Palermo G, Joris H, Devroey P, Van Steirteghem AC. Pregnancies after intracytoplasmic injection of single spermatozoon into an oocyte. *Lancet* 1992;340:17–8.
7. Rienzi L, Gracia C, Maggiulli R, LaBarbera AR, Kaser DJ, Ubaldi FM, et al. Oocyte, embryo and blastocyst cryopreservation in ART: systematic review and meta-analysis comparing slow-freezing versus vitrification to produce evidence for the development of global guidance. *Hum Reprod Update* 2017;23:139–55.
8. Ciani O, Buyse M, Drummond M, Rasi G, Saad ED, Taylor RS. Time to review the role of surrogate end points in health policy: state of the art and the way forward. *Value Health* 2017;20:487–95.
9. Manyara AM, Davies P, Stewart D, Weir CJ, Young A, Butcher NJ, et al. Protocol for the development of SPIRIT and CONSORT extensions for

- randomised controlled trials with surrogate primary endpoints: SPIRIT-SURROGATE and CONSORT-SURROGATE. *BMJ Open* 2022;12:e064304.
10. Grigore B, Ciani O, Dams F, Federici C, de Groot S, Möllenkamp M, et al. Surrogate endpoints in health technology assessment: an international review of methodological guidelines. *Pharmacoeconomics* 2020;38:1055–70.
11. De Gruttola VG, Clax P, DeMets DL, Downing GJ, Ellenberg SS, Friedman L, et al. Considerations in the evaluation of surrogate endpoints in clinical trials. Summary of a National Institutes of Health workshop. *Control Clin Trials* 2001;22:485–502.
12. Vaiarelli A, Zacà C, Spadoni V, Cimadomo D, Conforti A, Alviggi C, et al. Clinical and laboratory key performance indicators in IVF: a consensus between the Italian Society of Fertility and Sterility and Reproductive Medicine (SIFES-MR) and the Italian Society of Embryology, Reproduction and Research (SIERR). *J Assist Reprod Genet* 2023;40:1479–94.
13. Zacà C, Coticchio G, Vigiliano V, Lagalla C, Nadalini M, Tarozzi N, et al. Fine-tuning IVF laboratory key performance indicators of the Vienna consensus according to female age. *J Assist Reprod Genet* 2022;39:945–52.
14. Fabozzi G, Albricci L, Cimadomo D, Amendola MG, Sanges F, Maggiulli R, et al. Blastulation rates of sibling oocytes in two IVF culture media: an evidence-based workflow to implement newly commercialized products. *Reprod Biomed Online* 2021;42:311–22.
15. Wang HT, Hong PP, Li HY, Zhou W, Li T. Use of a new set of key performance indicators for evaluating the performance of an in vitro fertilization laboratory in which blastocyst culture and the freeze-all strategy are the primary treatment in patients with in vitro fertilization. *J Int Med Res* 2021;49:3000605211044364.
16. Fabozzi G, Cimadomo D, Maggiulli R, Vaiarelli A, Ubaldi FM, Rienzi L. Which key performance indicators are most effective in evaluating and managing an in vitro fertilization laboratory? *Fertil Steril* 2020;114:9–15.
17. Franco Jr JG, Petersen CG, Mauri AL, Vagnini LD, Renzi A, Petersen B, et al. Key performance indicators score (KPIs-score) based on clinical and laboratory parameters can establish benchmarks for internal quality control in an ART program. *JBRA Assist Reprod* 2017;21:61–6.
18. Piantadosi S. Clinical trials: a methodologic perspective. 2nd ed. Hoboken: Wiley; 2005:1–687.
19. Harper J, Magli MC, Lundin K, Barratt CL, Brison D. When and how should new technology be introduced into the IVF laboratory? *Hum Reprod* 2012;27:303–13.
20. Mortimer ST, Mortimer D. Quality and risk management in the IVF laboratory. 2nd edition. Cambridge, UK: Cambridge University Press; 2015.
21. Mortimer D, Pool TR, Cohen J. Introduction to quality management in assisted reproductive technology symposium. *Reprod Biomed Online* 2014;28:533–4.
22. Mortimer D, Mortimer ST. Quality and risk management in the IVF laboratory. Cambridge, UK: Cambridge University Press; 2005.
23. Mortimer D. Quality management in the IVF laboratory. In: Jansen R, Mortimer D, editors. Towards reproductive certainty: fertility and genetics beyond 1999: the plenary proceedings of the 11th World Congress on In Vitro Fertilization & Human Reproductive Genetics. Pearl River, NY: Parthenon; 1999.
24. Fischer C, Scott Jr RT. Three simple metrics to define in vitro fertilization success rates. *Fertil Steril* 2020;114:6–8.
25. Agarwal A, Sharma RK. Automation is the key to standardized semen analysis using the automated SQA-V sperm quality analyzer. *Fertil Steril* 2007;87:156–62.
26. Sheibak N, Amjadi F, Shamloo A, Zarei F, Zandieh Z. Microfluidic sperm sorting selects a subpopulation of high-quality sperm with a higher potential for fertilization. *Hum Reprod* 2024;39:902–11.
27. Roy TK, Brandi S, Tappe NM, Bradley CK, Vom E, Henderson C, et al. Embryo vitrification using a novel semi-automated closed system yields in vitro outcomes equivalent to the manual Cryotop method. *Hum Reprod* 2014;29:2431–8.
28. Wong CC, Loewke KE, Bossert NL, Behr B, De Jonge CJ, Baer TM, et al. Non-invasive imaging of human embryos before embryonic genome activation predicts development to the blastocyst stage. *Nat Biotechnol* 2010;28:1115–21.
29. Zou H, Wang R, Morbeck DE. Diagnostic or prognostic? Decoding the role of embryo selection on in vitro fertilization treatment outcomes. *Fertil Steril* 2024;121:730–6.
30. Cimadomo D, Soscia D, Vaiarelli A, Maggiulli R, Capalbo A, Ubaldi FM, et al. Looking past the appearance: a comprehensive description of the clinical contribution of poor-quality blastocysts to increase live birth rates during cycles with aneuploidy testing. *Hum Reprod* 2019;34:1206–14.
31. Corti L, Cermisoni GC, Alteri A, Pagliardini L, Ambrosini G, Andrisani A, et al. Clinical outcomes deriving from transfer of blastocysts developed in day 7: a systematic review and meta-analysis of frozen-thawed IVF cycles. *Reprod Sci* 2022;29:43–53.
32. ESHRE Special Interest Group of Embryology; Alpha Scientists in Reproductive Medicine. The Vienna consensus: report of an expert meeting on the development of art laboratory performance indicators. *Hum Reprod Open* 2017;2017:hox011.
33. ESHRE Special Interest Group of Embryology and Alpha Scientists in Reproductive Medicine. The Vienna consensus: report of an expert meeting on the development of ART laboratory performance indicators. *Reprod Biomed Online* 2017;35:494–510.
34. Wirleitner B, Schuff M, Stecher A, Murtinger M, Vanderzwalmen P. Pregnancy and birth outcomes following fresh or vitrified embryo transfer according to blastocyst morphology and expansion stage, and culturing strategy for delayed development. *Hum Reprod* 2016;31:1685.
35. Gardner DK, Schoolcraft WB. Culture and transfer of human blastocysts. *Curr Opin Obstet Gynecol* 1999;11:307–11.
36. Hammond ER, Morbeck DE. Day 5 useable blastocyst rate is an early indicator of clinically significant shifts in the embryo culture system. *Fertil Steril* 2018;110:e358–9.
37. Storr A, Venetis CA, Cooke S, Kilani S, Ledger W. Inter-observer and intra-observer agreement between embryologists during selection of a single day 5 embryo for transfer: a multicenter study. *Hum Reprod* 2017;32:307.
38. Cimadomo D, Forman EJ, Morbeck DE, Liperis G, Miller K, Zaninovic N, et al. Day7 and low-quality blastocysts: opt in or opt out? A dilemma with important clinical implications. *Fertil Steril* 2023;120:1151–9.
39. Scott L, Finn A, O'Leary T, McLellan S, Hill J. Morphologic parameters of early cleavage-stage embryos that correlate with fetal development and delivery: prospective and applied data for increased pregnancy rates. *Hum Reprod* 2007;22:230–40.
40. Ciray HN, Karagenc L, Ulug U, Bener F, Bahceci M. Early cleavage morphology affects the quality and implantation potential of day 3 embryos. *Fertil Steril* 2006;85:358–65.
41. Gardner DK, Sakkas D. Assessment of embryo viability: the ability to select a single embryo for transfer—a review. *Placenta* 2003;24:55–12.
42. Lundin K, Bergh C, Hardarson T. Early embryo cleavage is a strong indicator of embryo quality in human IVF. *Hum Reprod* 2001;16:2652–7.
43. Scott L, Alvero R, Leondires M, Miller B. The morphology of human pronuclear embryos is positively related to blastocyst development and implantation. *Hum Reprod* 2000;15:2394–403.
44. Racowsky C, Stern JE, Gibbons WE, Behr B, Pomeroy KO, Biggers JD. National collection of embryo morphology data into Society for Assisted Reproductive Technology Clinic Outcomes Reporting System: associations among day 3 cell number, fragmentation and blastomere asymmetry, and live birth rate. *Fertil Steril* 2011;95:1985–9.
45. Asami M, Lam BYH, Ma MK, Rainbow K, Braun S, VerMilyea MD, et al. Human embryonic genome activation initiates at the one-cell stage. *Cell Stem Cell* 2022;29:209–16.e4.
46. Shen X, Long H, Gao H, Guo W, Xie Y, Chen D, et al. The valuable reference of live birth rate in the single vitrified-warmed BB/BC/CB blastocyst transfer: the cleavage-stage embryo quality and embryo development speed. *Front Physiol* 2020;11:1102.
47. Wu J, Zhang J, Kuang Y, Chen Q, Wang Y. The effect of day 3 cell number on pregnancy outcomes in vitrified-thawed single blastocyst transfer cycles. *Hum Reprod* 2020;35:2478–87.
48. Shebl O, Haslinger C, Kresic S, Enengl S, Reiter E, Oppelt P, et al. The hare and the tortoise: extreme mitotic rates and how these affect live birth. *Reprod Biomed Online* 2021;42:332–9.

49. Lane SL, Reed L, Schoolcraft WB, Katz-Jaffe MG. Euploid day 7 blastocysts of infertility patients with only slow embryo development have reduced implantation potential. *Reprod Biomed Online* 2022;44:858–65.
50. Abdala A, Elkhatib I, Bayram A, Armanz A, El-Damen A, Melado L, et al. Day 5 vs day 6 single euploid blastocyst frozen embryo transfers: which variables do have an impact on the clinical pregnancy rates? *J Assist Reprod Genet* 2022;39:379–88.
51. Liu X, Lou H, Zhang J, Du M, Du Y, Wu S, et al. Clinical outcome analysis of frozen-thawed embryo transfer on day 7. *Front Endocrinol (Lausanne)* 2022;13:1082597.
52. Bourdon M, Pocate-Cheriet K, Finet De Bantel A, Grzegorzczak-Martin V, Amar Hoffer A, Arbo E, et al. Day 5 versus day 6 blastocyst transfers: a systematic review and meta-analysis of clinical outcomes. *Hum Reprod* 2019;34:1948–64.
53. Hammond ER, Cree LM, Morbeck DE. Should extended blastocyst culture include day 7? *Hum Reprod* 2018;33:991–7.
54. Ahlström A, Westin C, Reisner E, Wikland M, Hardarson T. Trophoctoderm morphology: an important parameter for predicting live birth after single blastocyst transfer. *Hum Reprod* 2011;26:3289.
55. Bakkensen JB, Brady P, Carusi D, Romanski P, Thomas AM, Racowsky C. Association between blastocyst morphology and pregnancy and perinatal outcomes following fresh and cryopreserved embryo transfer. *J Assist Reprod Genet* 2019;36:2315–24.
56. Pons MC, Carrasco B, Rives N, Delgado A, Martínez-Moro A, Martínez-Granados L, et al. Predicting the likelihood of live birth: an objective and user-friendly blastocyst grading system. *Reprod Biomed Online* 2023;47:103243.
57. Thompson SM, Onwubalili N, Brown K, Jindal SK, McGovern PG. Blastocyst expansion score and trophoctoderm morphology strongly predict successful clinical pregnancy and live birth following elective single embryo blastocyst transfer (eSET): a national study. *J Assist Reprod Genet* 2013;30:1577–81.
58. Du QY, Wang EY, Huang Y, Guo XY, Xiong YJ, Yu YP, et al. Blastocoele expansion degree predicts live birth after single blastocyst transfer for fresh and vitrified/warmed single blastocyst transfer cycles. *Fertil Steril* 2016;105:910–9.e1.
59. Ai J, Jin L, Zheng Y, Yang P, Huang B, Dong X. The morphology of inner cell mass is the strongest predictor of live birth after a frozen-thawed single embryo transfer. *Front Endocrinol (Lausanne)* 2021;12:621221.
60. Subira J, Craig J, Turner K, Bevan A, Ohuma E, McVeigh E, et al. Grade of the inner cell mass, but not trophoctoderm, predicts live birth in fresh blastocyst single transfers. *Hum Fertil (Camb)* 2016;19:254–61.
61. Gardner DK, Lane M, Stevens J, Schlenker T, Schoolcraft WB. Blastocyst score affects implantation and pregnancy outcome: towards a single blastocyst transfer. *Fertil Steril* 2000;73:1155–8.
62. Bormann CL, Curchoe CL, Thirumalaraju P, Kanakasabapathy MK, Gupta R, Pooniwal R, et al. Deep learning early warning system for embryo culture conditions and embryologist performance in the ART laboratory. *J Assist Reprod Genet* 2021;38:1641–6.
63. Chavez-Badiola A, Flores-Saiffe-Farias A, Mendizabal-Ruiz G, Drakeley AJ, Cohen J. Embryo Ranking Intelligent Classification Algorithm (ERICA): artificial intelligence clinical assistant predicting embryo ploidy and implantation. *Reprod Biomed Online* 2020;41:585–93.
64. Cimadomo D, Soscia D, Casciani V, Innocenti F, Trio S, Chiappetta V, et al. How slow is too slow? A comprehensive portrait of Day 7 blastocysts and their clinical value standardized through artificial intelligence. *Hum Reprod* 2022;37:1134–47.
65. Diakiw SM, Hall JMM, VerMilyea M, Lim AYY, Quangkananurug W, Chanchamroen S, et al. An artificial intelligence model correlated with morphological and genetic features of blastocyst quality improves ranking of viable embryos. *Reprod Biomed Online* 2022;45:1105–17.
66. Diakiw SM, Hall JMM, VerMilyea MD, Amin J, Aizpurua J, Giardini L, et al. Development of an artificial intelligence model for predicting the likelihood of human embryo euploidy based on blastocyst images from multiple imaging systems during IVF. *Hum Reprod* 2022;37:1746–59.
67. Huang TTF, Kosasa T, Walker B, Arnett C, Huang CTF, Yin C, et al. Deep learning neural network analysis of human blastocyst expansion from time-lapse image files. *Reprod Biomed Online* 2021;42:1075–85.
68. Khosravi P, Kazemi E, Zhan Q, Malmsten JE, Toschi M, Zisimopoulos P, et al. Deep learning enables robust assessment and selection of human blastocysts after in vitro fertilization. *NPJ Digit Med* 2019;2:21.
69. Loewke K, Cho JH, Brumar CD, Maeder-York P, Barash O, Malmsten JE, et al. Characterization of an artificial intelligence model for ranking static images of blastocyst stage embryos. *Fertil Steril* 2022;117:528–35.
70. VerMilyea M, Miller A, Lane M, Adaniya G, Bopp B, Morbeck DE, et al. Artificial intelligence (AI) technology can predict human embryo viability across multiple laboratories with varying demographics with high accuracy and reproducibility. *Hum Reprod* 2019;34:2.
71. Imudia AN, Kumar S, Diamond MP, DeCherney AH, Armant DR. Transcervical retrieval of fetal cells in the practice of modern medicine: a review of the current literature and future direction. *Fertil Steril* 2010;93:1725–30.
72. Fritz R, Kohan-Ghadr HR, Bolnick JM, Bolnick AD, Kilburn BA, Diamond MP, et al. Noninvasive detection of trophoblast protein signatures linked to early pregnancy loss using trophoblast retrieval and isolation from the cervix (TRIC). *Fertil Steril* 2015;104:339–46.e4.
73. Fritz R, Bolnick J, Bolnick A, Modi M, Kilburn B, Diamond MP, et al. Trophoblast retrieval and isolation from the cervix (TRIC) to predict risk for spontaneous abortion. *Fertil Steril* 2014;102:e55–6.
74. Campbell A, Fishel S, Bowman N, Duffy S, Sedler M, Hickman CF. Modeling a risk classification of aneuploidy in human embryos using non-invasive morphokinetics. *Reprod Biomed Online* 2013;26:477–85.
75. Fishel S, Campbell A, Foad F, Davies L, Best L, Davis N, et al. Evolution of embryo selection for IVF from subjective morphology assessment to objective time-lapse algorithms improves chance of live birth. *Reprod Biomed Online* 2020;40:61–70.
76. Bamford T, Smith R, Young S, Evans A, Lockwood M, Easter C, et al. A comparison of morphokinetic models and morphological selection for prioritizing euploid embryos: a multicentre cohort study. *Hum Reprod* 2024;39:53–61.
77. Capalbo A, Rienzi L, Cimadomo D, Maggiulli R, Elliott T, Wright G, et al. Correlation between standard blastocyst morphology, euploidy and implantation: an observational study in two centers involving 956 screened blastocysts. *Hum Reprod* 2014;29:1173–81.
78. Tiegs AW, Sun L, Patounakis G, Scott RT. Worth the wait? Day 7 blastocysts have lower euploidy rates but similar sustained implantation rates as day 5 and day 6 blastocysts. *Hum Reprod* 2019;34:1632–9.
79. Zhan Q, Sierra ET, Malmsten J, Ye Z, Rosenwaks Z, Zaninovic N. Blastocyst score, a blastocyst quality ranking tool, is a predictor of blastocyst ploidy and implantation potential. *F S Rep* 2020;1:133–41.
80. Irani M, Reichman D, Robles A, Melnick A, Davis O, Zaninovic N, et al. Morphologic grading of euploid blastocysts influences implantation and ongoing pregnancy rates. *Fertil Steril* 2017;107:664–70.
81. Munné S, Alikani M, Ribustello L, Colls P, Martínez-Ortiz PA, McCulloh DH. Euploidy rates in donor egg cycles significantly differ between fertility centers. *Hum Reprod* 2017;32:743–9.
82. Abdala A, Elkhatib I, Bayram A, Armanz A, El-Damen A, Melado L, et al. Different CO2 settings (6.0% vs 7.0%) do have an impact on extracellular pH of culture medium (pHe) and euploidy rates rather than on blastocyst development: a sibling oocyte study. *J Assist Reprod Genet* 2021;38:2915–23.
83. Tiegs AW, Tao X, Zhan Y, Whitehead C, Kim J, Hanson B, et al. A multicenter, prospective, blinded, nonselection study evaluating the predictive value of an aneuploid diagnosis using a targeted next-generation sequencing-based preimplantation genetic testing for aneuploidy assay and impact of biopsy. *Fertil Steril* 2021;115:627–37.
84. Wang Y, Taylor JM. A measure of the proportion of treatment effect explained by a surrogate marker. *Biometrics* 2002;58:803–12.
85. Zhuang R, Chen YQ. Measuring surrogacy in clinical research: with an application to studying surrogate markers for HIV treatment-as-prevention. *Stat Biosci* 2020;12:295–323.
86. Agniel D, Parast L. Evaluation of longitudinal surrogate markers. *Biometrics* 2021;77:477–89.
87. Schellekens H. Biosimilar therapeutics-what do we need to consider? *NDT Plus* 2009;2:i27–36.
88. Epstein M. Food and Drug Administration guidances on biosimilars: an update for the gastroenterologist. *Ther Adv Gastroenterol* 2018;11:1756284818799600.

89. Maheshwari A, McLernon D, Bhattacharya S. Cumulative live birth rate: time for a consensus? *Hum Reprod* 2015;30:2703–7.
90. Bartolacci A, Tondo F, Alteri A, Solano Narduche L, de Girolamo S, D'Alessandro G, et al. The task matters: a comprehensive review and proposed literature score of the effects of chemical and physical parameters on embryo developmental competence. *Life (Basel)* 2023;13:2161.
91. Higdon HL 3rd, Blackhurst DW, Boone WR. Incubator management in an assisted reproductive technology laboratory. *Fertil Steril* 2008;89:703–10.
92. Wang WH, Meng L, Hackett RJ, Oldenbourg R, Keefe DL. Rigorous thermal control during intracytoplasmic sperm injection stabilizes the meiotic spindle and improves fertilization and pregnancy rates. *Fertil Steril* 2002;77:1274–7.
93. Hong KH, Lee H, Forman EJ, Upham KM, Scott Jr RT. Examining the temperature of embryo culture in vitro fertilization: a randomized controlled trial comparing traditional core temperature (37°C) to a more physiologic, cooler temperature (36°C). *Fertil Steril* 2014;102:767–73.
94. Neelke M, Ronny J, Samuel SR, Herman T, Hilde VV, Greta V. The effect of different temperature conditions on human embryos in vitro: two sibling studies. *Reprod Biomed Online* 2019;38:508–15.
95. Fawzy M, Emad M, Gad MA, Sabry M, Kasem H, Mahmoud M, et al. Comparing 36.5°C with 37°C for human embryo culture: a prospective randomized controlled trial. *Reprod Biomed Online* 2018;36:620–6.
96. Dumoulin JC, Vanvuchelen RC, Land JA, Pieters MH, Geraedts JP, Evers JL. Effect of oxygen concentration on in vitro fertilization and embryo culture in the human and the mouse. *Fertil Steril* 1995;63:115–9.
97. Dumoulin JC, Meijers CJ, Bras M, Coonen E, Geraedts JP, Evers JL. Effect of oxygen concentration on human in-vitro fertilization and embryo culture. *Hum Reprod* 1999;14:465–9.
98. Kea B, Gebhardt J, Watt J, Westphal LM, Lathi RB, Milki AA, et al. Effect of reduced oxygen concentrations on the outcome of in vitro fertilization. *Fertil Steril* 2007;87:213–6.
99. Ciray HN, Aksoy T, Yaramanci K, Karayaka I, Bahceci M. In vitro culture under physiologic oxygen concentration improves blastocyst yield and quality: a prospective randomized survey on sibling oocytes. *Fertil Steril* 2009;91:1459–61.
100. Waldenström U, Engström AB, Hellberg D, Nilsson S. Low-oxygen compared with high-oxygen atmosphere in blastocyst culture, a prospective randomized study. *Fertil Steril* 2009;91:2461–5.
101. Kovacic B, Vlaisavljevic V. Influence of atmospheric versus reduced oxygen concentration on development of human blastocysts in vitro: a prospective study on sibling oocytes. *Reprod Biomed Online* 2008;17:229–36.
102. Meintjes M, Chantilis SJ, Douglas JD, Rodriguez AJ, Guerami AR, Bookout DM, et al. A controlled randomized trial evaluating the effect of lowered incubator oxygen tension on live births in a predominantly blastocyst transfer program. *Hum Reprod* 2009;24:300–7.
103. Kovacic B, Sajko MC, Vlaisavljevic V. A prospective, randomized trial on the effect of atmospheric versus reduced oxygen concentration on the outcome of intracytoplasmic sperm injection cycles. *Fertil Steril* 2010;94:511–9.
104. Guo N, Li Y, Ai J, Gu L, Chen W, Liu Q. Two different concentrations of oxygen for culturing precompaction stage embryos on human embryo development competence: a prospective randomized sibling-oocyte study. *Int J Clin Exp Pathol* 2014;7:6191–8.
105. Yang Y, Xu Y, Ding C, khoudja RY, Lin M, Awonuga AO, et al. Comparison of 2, 5, and 20 % O₂ on the development of post-thaw human embryos. *J Assist Reprod Genet* 2016;33:919–27.
106. Ferrieres-Hoa AF, Gala A, Haouzi D, Roman K, Hamamah S. Ultra-low (2%) oxygen tension positively affects blastocyst quality. *Fertil Steril* 2017;108:e151.
107. Kaser DJ, Bogale B, Sarda V, Farland LV, Williams PL, Racowsky C. Randomized controlled trial of low (5%) versus ultralow (2%) oxygen for extended culture using bipronucleate and tripronucleate human preimplantation embryos. *Fertil Steril* 2018;109:1030–7.e2.
108. Li M, Xue X, Shi J. Ultralow oxygen tension (2%) is beneficial for blastocyst formation of in vitro human low-quality embryo culture. *Biomed Res Int* 2022;2022:9603185.
109. Papadopoulou MI, Karagianni M, Vorniotaki A, Oraiopoulou C, Christophoridis N, Papatheodorou A, et al. P-169 Low 5% VS ultra-low 3% O₂ concentration on embryo culture: is there any difference in quality and ploidy? *Hum Reprod* 2022;37.
110. Chen HH, Lee CI, Huang CC, Cheng EH, Lee TH, Lin PY, et al. Biphasic oxygen tension promotes the formation of transferable blastocysts in patients without euploid embryos in previous monophasic oxygen cycles. *Sci Rep* 2023;13:4330.
111. De Munck N, Janssens R, Segers I, Tournaye H, Van de Velde H, Verheyen G. Influence of ultra-low oxygen (2%) tension on in-vitro human embryo development. *Hum Reprod* 2019;34:228–34.
112. Brouillet S, Baron C, Barry F, Andreeva A, Haouzi D, Gala A, et al. Biphasic (5–2%) oxygen concentration strategy significantly improves the usable blastocyst and cumulative live birth rates in in vitro fertilization. *Sci Rep* 2021;11:22461.
113. Herbemont C, Labrosse J, Bennani-Smires B, Cedrin-Dumerin I, Peigne M, Sermondade N, et al. Impact of oxygen tension according to embryo stage of development: a prospective randomized study. *Sci Rep* 2021;11:22313.
114. Fawzy M, AbdelRahman MY, Zidan MH, Abdel Hafez FF, Abdelghafar H, Al-Inany H, et al. Humid versus dry incubator: a prospective, randomized, controlled trial. *Fertil Steril* 2017;108:277–83.
115. Swain JE, Graham C, Kile R, Schoolcraft WB, Krisher RL. Media evaporation in a dry culture incubator; effect of dish, drop size and oil on media osmolality. *Fertil Steril* 2018;110:e363–4.
116. Del Gallego R, Albert C, Marcos J, Larreategui Z, Alegre L, Meseguer M. Humid vs. dry embryo culture conditions on embryo development: a continuous embryo monitoring assessment. *Fertil Steril* 2018;110:e362–3.
117. Valera MÁ, Albert C, Marcos J, Larreategui Z, Bori L, Meseguer M. A propensity score-based, comparative study assessing humid and dry time-lapse incubation, with single-step medium, on embryo development and clinical outcomes. *Hum Reprod* 2022;37:1980–93.
118. Bartolacci A, Borini A, Cimadomo D, Fabozzi G, Maggiulli R, Lagalla C, et al. Humidified atmosphere in a time-lapse embryo culture system does not improve ongoing pregnancy rate: a retrospective propensity score model study derived from 496 first ICSI cycles. *J Assist Reprod Genet* 2023;40:1429–35.
119. Kirkegaard K, Ahlstrom A, Ingerslev HJ, Hardarson T. Choosing the best embryo by time lapse versus standard morphology. *Fertil Steril* 2015;103:323–32.
120. Sciorio R, Thong JK, Pickering SJ. Comparison of the development of human embryos cultured in either an EmbryoScope or benchtop incubator. *J Assist Reprod Genet* 2018;35:515–22.
121. Zhang XD, Zhang Q, Han W, Liu WW, Shen XL, Yao GD, et al. Comparison of embryo implantation potential between time-lapse incubators and standard incubators: a randomized controlled study. *Reprod Biomed Online* 2022;45:858–66.
122. Nobrega NG, Abdala A, El-Damen A, Arnanz A, Bayram A, Elkhatab I, et al. Sibling oocytes cultured in a time-lapse versus benchtop incubator: how time-lapse incubators improve blastocyst development and euploid rate. *Zygote* 2023;18:402–9.
123. Cruz M, Gadea B, Garrido N, Pedersen KS, Martinez M, Perez-Cano I, et al. Embryo quality, blastocyst and ongoing pregnancy rates in oocyte donation patients whose embryos were monitored by time-lapse imaging. *J Assist Reprod Genet* 2011;28:569–73.
124. Barberet J, Chammass J, Bruno C, Valot E, Vuillemin C, Jonval L, et al. Randomized controlled trial comparing embryo culture in two incubator systems: G185 K-System versus EmbryoScope. *Fertil Steril* 2018;109:302–9.e1.
125. Kalleas D, McEvoy K, Horne G, Roberts SA, Brison DR. Live birth rate following undisturbed embryo culture at low oxygen in a time-lapse incubator compared to a high-quality benchtop incubator. *Hum Fertil (Camb)* 2022;25:147–53.
126. Swain JE. Optimal human embryo culture. *Semin Reprod Med* 2015;33:103–17.
127. Swain JE. Decisions for the IVF laboratory: comparative analysis of embryo culture incubators. *Reprod Biomed Online* 2014;28:535–47.

128. Bódis J, Gödöny K, Várnagy Á, Kovács K, Koppán M, Nagy B, et al. How to reduce the potential harmful effects of light on blastocyst development during IVF. *Med Princ Pract* 2020;29:558–64.
129. Hentemann M, Mousavi K, Bertheussen K. Differential pH in embryo culture. *Fertil Steril* 2011;95:1291–4.
130. Bontekoe S, Mantikou E, van Wely M, Seshadri S, Repping S, Mastenbroek S. Low oxygen concentrations for embryo culture in assisted reproductive technologies. *Cochrane Database Syst Rev* 2012;2012:CD008950.
131. Meintjes M, Chantilis SJ, Ward DC, Douglas JD, Rodríguez AJ, Guerami AR, et al. A randomized controlled study of human serum albumin and serum substitute supplement as protein supplements for IVF culture and the effect on live birth rates. *Hum Reprod* 2009;24:782–9.
132. Ruiz M, Santamaría-López E, Blasco V, Hernández MJ, Caligara C, Pellicer A, et al. Effect of group embryo culture under low-oxygen tension in bench-top incubators on human embryo culture: prospective, randomized, controlled trial. *Reprod Sci* 2020;27:1522–33.
133. Glatthorn HN, Hanson BM, Kim JG, Herlihy NS, Klimczak AM, Hong KH, et al. Individual culture leads to decreased blastocyst formation but does not affect pregnancy outcomes in the setting of a single, vitrified-warmed euploid blastocyst transfer. *J Assist Reprod Genet* 2021;38:2157–64.
134. Hong KH, Forman EJ, Lee H, Upham KM, Treff NR, Scott RT. Optimizing the temperature of embryo culture in IVF: a randomized controlled trial. *Fertil Steril* 2013;100:S26.
135. Morbeck DE, Baumann NA, Oglesbee D. Composition of single-step media used for human embryo culture. *Fertil Steril* 2017;107:1055–60.e1.
136. Morbeck DE, Krisher RL, Herrick JR, Baumann NA, Matern D, Moyer T. Composition of commercial media used for human embryo culture. *Fertil Steril* 2014;102:759–66.e9.
137. Morbeck DE, Paczkowski M, Fredrickson JR, Krisher RL, Hoff HS, Baumann NA, et al. Composition of protein supplements used for human embryo culture. *J Assist Reprod Genet* 2014;31:1703–11.
138. Youssef MM, Mantikou E, van Wely M, Van der Veen F, Al-Inany HG, Repping S, et al. Culture media for human pre-implantation embryos in assisted reproductive technology cycles. *Cochrane Database Syst Rev* 2015;2015:CD007876.
139. Legro RS, Diamond MP, Coutifaris C, Schlaff WD, Alvero R, Casson P, et al. Pregnancy registry: three-year follow-up of children conceived from letrozole, clomiphene, or gonadotropins. *Fertil Steril* 2020;113:1005–13.
140. Penova-Veselinovic B, Wijs LA, Yovich JL, Burton P, Hart RJ. Cohort profile: the Growing Up Healthy Study (GUHS)-a prospective and observational cohort study investigating the long-term health outcomes of offspring conceived after assisted reproductive technologies. *PLOS One* 2022;17:e0272064.
141. Ono M, Kuji N, Ueno K, Kojima J, Nishi H. The long-term outcome of children conceived through assisted reproductive technology. *Reprod Sci* 2024;31:583–90.
142. Belbasis L, Savvidou MD, Kanu C, Evangelou E, Tzoulaki I. Birth weight in relation to health and disease in later life: an umbrella review of systematic reviews and meta-analyses. *BMC Med* 2016;14:147.
143. Jańczewska I, Wierzbą J, Jańczewska A, Szczurek-Gierczak M, Domżańska-Popadiuk I. Prematurity and low birth weight and their impact on childhood growth patterns and the risk of long-term cardiovascular sequelae. *Children (Basel)* 2023;10:1599.
144. Pravia CI, Benny M. Long-term consequences of prematurity. *Cleve Clin J Med* 2020;87:759–67.
145. Scifres CM. Short- and long-term outcomes associated with large for gestational age birth weight. *Obstet Gynecol Clin North Am* 2021;48:325–37.
146. Berntsen S, Söderström-Anttila V, Wennerholm UB, Laivuori H, Loft A, Oldereid NB, et al. The health of children conceived by ART: 'the chicken or the egg?'. *Hum Reprod Update* 2019;25:137–58.
147. Elias FTS, Weber-Adrian D, Pudwell J, Carter J, Walker M, Gaudet L, et al. Neonatal outcomes in singleton pregnancies conceived by fresh or frozen embryo transfer compared to spontaneous conceptions: a systematic review and meta-analysis. *Arch Gynecol Obstet* 2020;302:31–45.
148. Mol BW, Jacobsson B, Grobman WA, Moley K, FIGO Working Group for Preterm Birth. FIGO good practice recommendations on reduction of preterm birth in pregnancies conceived by assisted reproductive technologies. *Int J Gynaecol Obstet* 2021;155:13–5.
149. Stern JE, Farland LV, Hwang SS, Dukhovny D, Coddington CC, Cabral HJ, et al. Assisted reproductive technology or infertility: what underlies adverse outcomes? Lessons from the Massachusetts Outcome Study of Assisted Reproductive Technology. *F S Rev* 2022;3:242–55.
150. Pinborg A, Wennerholm UB, Romundstad LB, Loft A, Aittomäki K, Söderström-Anttila V, et al. Why do singletons conceived after assisted reproduction technology have adverse perinatal outcome? Systematic review and meta-analysis. *Hum Reprod Update* 2013;19:87–104.
151. Magnusson Å, Laivuori H, Loft A, Oldereid NB, Pinborg A, Petzold M, et al. The association between high birth weight and long-term outcomes-implications for assisted reproductive technologies: a systematic review and meta-analysis. *Front Pediatr* 2021;9:675775.
152. Davies MJ, Moore VM, Willson KJ, Van Essen P, Priest K, Scott H, et al. Reproductive technologies and the risk of birth defects. *N Engl J Med* 2012;366:1803–13.
153. Hansen M, Kurinczuk JJ, Milne E, de Klerk N, Bower C. Assisted reproductive technology and birth defects: a systematic review and meta-analysis. *Hum Reprod Update* 2013;19:330–53.
154. Zhu JL, Basso O, Obel C, Bille C, Olsen J. Infertility, infertility treatment, and congenital malformations: Danish national birth cohort. *BMJ* 2006;333:679–81.
155. Bergh C, Wennerholm UB. Obstetric outcome and long-term follow up of children conceived through assisted reproduction. *Best Pract Res Clin Obstet Gynaecol* 2012;26:841–52.
156. Graham ME, Jelin A, Hoon Jr AH, Wilms Floet AM, Levey E, Graham EM. Assisted reproductive technology: short- and long-term outcomes. *Dev Med Child Neurol* 2023;65:38–49.
157. Kleijkers SH, Mantikou E, Slappendel E, Consten D, van Echten-Arends J, Wetzels AM, et al. Influence of embryo culture medium (G5 and HTF) on pregnancy and perinatal outcome after IVF: a multicenter RCT. *Hum Reprod* 2016;31:2219–30.
158. Siristatidis C, Papapanou M, Karageorgiou V, Martins WP, Bellos I, Teixeira DM, et al. Congenital anomaly and perinatal outcome following blastocyst- vs cleavage-stage embryo transfer: systematic review and network meta-analysis. *Ultrasound Obstet Gynecol* 2023;61:12–25.
159. Castillo CM, Harper J, Roberts SA, O'Neill HC, Johnstone ED, Brison DR. The impact of selected embryo culture conditions on ART treatment cycle outcomes: a UK national study. *Hum Reprod Open* 2020;2020:hoz031.
160. Zandstra H, Van Montfoort AP, Dumoulin JC. Does the type of culture medium used influence birthweight of children born after IVF? *Hum Reprod* 2015;30:2693.
161. Dumoulin JC, Land JA, Van Montfoort AP, Nelissen EC, Coonen E, Derhaag JG, et al. Effect of in vitro culture of human embryos on birthweight of newborns. *Hum Reprod* 2010;25:605–12.
162. Ginström Ernstad E, Wennerholm UB, Khatibi A, Petzold M, Bergh C. Neonatal and maternal outcome after frozen embryo transfer: Increased risks in programmed cycles. *Am J Obstet Gynecol* 2019;221:126.e1–18.
163. Pier BD, Roshong A, Santoro N, Sammel MD. Association of duration of embryo culture with risk of large for gestational age delivery in cryopreserved embryo transfer cycles. *Fertil Steril* 2024;121:814–23.
164. Maheshwari A, Hamilton M, Bhattacharya S. Should we be promoting embryo transfer at blastocyst stage? *Reprod Biomed Online* 2016;32:142–6.
165. Glujovsky D, Quinteiro Retamar AM, Alvarez Sedo CR, Ciapponi A, Cornelisse S, Blake D. Cleavage-stage versus blastocyst-stage embryo transfer in assisted reproductive technology. *Cochrane Database Syst Rev* 2022;2022:CD002118.
166. Marconi N, Raja EA, Bhattacharya S, Maheshwari A. Perinatal outcomes in singleton live births after blastocyst transfer: an analysis of 60,926 in vitro fertilization cycles from the United Kingdom. *Fertil Steril* 2023;120:312–20.
167. Shah JS, Vaughan DA, Leung A, Korkidakis A, Figueras F, Garcia D, et al. Perinatal outcomes in singleton pregnancies after in vitro fertilization cycles over 24 years. *Fertil Steril* 2021;116:27–35.

168. Maas K, Galkina E, Thornton K, Penzias AS, Sakkas D. No change in live birthweight of IVF singleton deliveries over an 18-year period despite significant clinical and laboratory changes. *Hum Reprod* 2016;31:1987–96.
169. Castillo CM, Horne G, Fitzgerald CT, Johnstone ED, Brison DR, Roberts SA. The impact of IVF on birthweight from 1991 to 2015: a cross-sectional study. *Hum Reprod* 2019;34:920–31.
170. Koeck RM, Busato F, Tost J, Zandstra H, Remy S, Langie S, et al. At age 9, the methylome of assisted reproductive technology children that underwent embryo culture in different media is not significantly different on a genome-wide scale. *Hum Reprod* 2022;37:2709–21.
171. Estill MS, Bolnick JM, Waterland RA, Bolnick AD, Diamond MP, Krawetz SA. Assisted reproductive technology alters deoxyribonucleic acid methylation profiles in bloodspots of newborn infants. *Fertil Steril* 2016;106:629–39.e10.
172. Duranthon V, Chavatte-Palmer P. Long term effects of ART: what do animals tell us? *Mol Reprod Dev* 2018;85:348–68.
173. Hanna CW, Demond H, Kelsey G. Epigenetic regulation in development: is the mouse a good model for the human? *Hum Reprod Update* 2018;24:556–76.
174. Henningsen AA, Gissler M, Rasmussen S, Opdahl S, Wennerholm UB, Spangsmose AL, et al. Imprinting disorders in children born after ART: a Nordic study from the CoNARTaS group. *Hum Reprod* 2020;35:1178–84.
175. Halliday J, Oke K, Breheny S, Algar E, J Amor D. Beckwith-Wiedemann syndrome and IVF: a case-control study. *Am J Hum Genet* 2004;75:526–8.
176. Walker AR, Venetis CA, Opdahl S, Chambers GM, Jorm LR, Vajdic CM. Estimating the impact of bias in causal epidemiological studies: the case of health outcomes following assisted reproduction. *Hum Reprod* 2024;39:869–75.
177. Christianson MS, Zhao Y, Shoham G, Granot I, Safran A, Khafagy A, et al. Embryo catheter loading and embryo culture techniques: results of a worldwide Web-based survey. *J Assist Reprod Genet* 2014;31:1029–36.
178. Rendón Abad M, Serra V, Gámiz P, de los Santos JM, Remohí J, Navarro AT, et al. The influence of oxygen concentration during embryo culture on obstetric and neonatal outcomes: a secondary analysis of a randomized controlled trial. *Hum Reprod* 2020;35:2017–25.
179. Munch EM, Sparks AE, Duran HE, Van Voorhis BJ. Lack of carbon air filtration impacts early embryo development. *J Assist Reprod Genet* 2015;32:1009–17.
180. Turner T. The identification of a toxic substance in the in vitro fertilization laboratory: the value of inter-laboratory communication. *Fertil Mag* 2010;12:64–5.
181. Wolff HS, Fredrickson JR, Walker DL, Morbeck DE. Advances in quality control: mouse embryo morphokinetics are sensitive markers of in vitro stress. *Hum Reprod* 2013;28:1776–82.
182. Adam MP. The all-or-none phenomenon revisited. *Birth Defects Res A Clin Mol Teratol* 2012;94:664–9.
183. Alwan S, Chambers CD. Identifying human teratogens: an update. *J Pediatr Genet* 2015;4:39–41.
184. Diamond MP, Willman S, Chenette P, Cedars MI. The clinical need for a method of identification of embryos destined to become a blastocyst in assisted reproductive technology cycles. *J Assist Reprod Genet* 2012;29:391–6.
185. Tarahomi M, Vaz FM, van Straalen JP, Schrauwen FAP, van Wely M, Hamer G, et al. The composition of human preimplantation embryo culture media and their stability during storage and culture. *Hum Reprod* 2019;34:1450–61.
186. Fawzy M, Emad M, Elsuity MA, Mahran A, Abdelrahman MY, Fetih AN, et al. Cytokines hold promise for human embryo culture in vitro: results of a randomized clinical trial. *Fertil Steril* 2019;112:849–57.e1.
187. Ziebe S, Loft A, Povlsen BB, Erb K, Agerholm I, Aasted M, et al. A randomized clinical trial to evaluate the effect of granulocyte-macrophage colony-stimulating factor (GM-CSF) in embryo culture medium for in vitro fertilization. *Fertil Steril* 2013;99:1600–9.e2.
188. Lemmen JG, Agerholm I, Ziebe S. Kinetic markers of human embryo quality using time-lapse recordings of IVF/ICSI-fertilized oocytes. *Reprod Biomed Online* 2008;17:385–91.
189. Mantikou E, Jonker MJ, Wong KM, van Montfoort AP, de Jong M, Breit TM, et al. Factors affecting the gene expression of in vitro cultured human pre-implantation embryos. *Hum Reprod* 2016;31:298–311.
190. Young LE, Sinclair KD, Wilmot I. Large offspring syndrome in cattle and sheep. *Rev Reprod* 1998;3:155–63.
191. Gardner DK, Kuramoto T, Tanaka M, Mizumoto S, Montag M, Yoshida A. Prospective randomized multicentre comparison on sibling oocytes comparing G-Series media system with antioxidants versus standard G-Series media system. *Reprod Biomed Online* 2020;40:637–44.
192. Prathalingam N, Hyslop L, Cole M, Cooney D, Driver A, Herbert M, et al. Developing a novel device, Eggcell, to improve temperature stability during oocyte collection for IVF. *Reprod Biomed Online* 2022;45:1097–104.
193. Venturas M, Yang X, Sakkas D, Needleman D. Noninvasive metabolic profiling of cumulus cells, oocytes, and embryos via fluorescence lifetime imaging microscopy: a mini-review. *Hum Reprod* 2023;38:799–810.
194. Zhai R, Shan G, Dai C, Hao M, Zhu J, Ru C, et al. Automated denudation of oocytes. *Micromachines (Basel)* 2022;13:1301.
195. Zhang Z, Moskovtsev S, Librach C, Jarvi K, Sun Y, Dai C, et al. Robotic immobilization of motile sperm for clinical intracytoplasmic sperm injection. *IEEE Trans Biomed Eng* 2019;66:444–52.
196. Costa-Borges N, Munné S, Albó E, Mas S, Castelló C, Giralt G, et al. First babies conceived with automated intracytoplasmic sperm injection. *Reprod Biomed Online* 2023;47:103237.
197. Arav A, Natan Y, Kalo D, Kornsky-Elbaz A, Roth Z, Levi-Setti PE, et al. A new, simple, automatic vitrification device: preliminary results with murine and bovine oocytes and embryos. *J Assist Reprod Genet* 2018;35:1161–8.
198. Morbeck DE. Importance of supply integrity for in vitro fertilization and embryo culture. *Semin Reprod Med* 2012;30:182–90.
199. Tucker N, Wright G, Morton P, Shanguo L, Massey J, Kort H. Preliminary experience with human oocyte cryopreservation using 1,2-propanediol and sucrose. *Hum Reprod* 1996;11:1513–5.
200. Chian RC, Son WY, Huang JY, Cui SJ, Buckett WM, Tan SL. High survival rates and pregnancies of human oocytes following vitrification: preliminary report. *Fertil Steril* 2005;84:S36.
201. Boldt J, Tidswell N, Sayers A, Kilani R, Cline D. Human oocyte cryopreservation: 5-year experience with a sodium-depleted slow freezing method. *Reprod Biomed Online* 2006;13:96–100.
202. Desai NN, Goldberg JM, Austin C, Falcone T. The new Rapid-i carrier is an effective system for human embryo vitrification at both the blastocyst and cleavage stage. *Reprod Biol Endocrinol* 2013;11:41.
203. Smith GD, Serafini PC, Fioravanti J, Yadid I, Coslovsky M, Hassun P, et al. Prospective randomized comparison of human oocyte cryopreservation with slow-rate freezing or vitrification. *Fertil Steril* 2010;94:2088–95.
204. Selman H, Angelini A, Barnocchi N, Brusco GF, Pacchiarotti A, Aragona C. Ongoing pregnancies after vitrification of human oocytes using a combined solution of ethylene glycol and dimethyl sulfoxide. *Fertil Steril* 2006;86:997–1000.
205. Rienzi L, Romano S, Albricci L, Maggiulli R, Capalbo A, Baroni E, et al. Embryo development of fresh 'versus' vitrified metaphase II oocytes after ICSI: a prospective randomized sibling-oocyte study. *Hum Reprod* 2010;25:66–73.
206. Guerrero J, Gallardo M, Rodríguez-Arnedo A, Ten J, Bernabeu R. Comparison of two closed carriers for vitrification of human blastocysts in a donor program. *Cryobiology* 2018;81:12–6.
207. Gallardo M, Hebles M, Migueles B, Dorado M, Aguilera L, González M, et al. Thermal and clinical performance of a closed device designed for human oocyte vitrification based on the optimization of the warming rate. *Cryobiology* 2016;73:40–6.
208. Gallardo M, Saenz J, Risco R. Human oocytes and zygotes are ready for ultra-fast vitrification after 2 minutes of exposure to standard CPA solutions. *Sci Rep* 2019;9:15986.
209. Kuleshova LL, Lopata A. Vitrification can be more favorable than slow cooling. *Fertil Steril* 2002;78:449–54.
210. Cimadomo D, Rienzi L, Conforti A, Forman E, Canosa S, Innocenti F, et al. Opening the black box: why do euploid blastocysts fail to

- implant? A systematic review and meta-analysis. *Hum Reprod Update* 2023;29:570–633.
211. Franasiak JM, Forman EJ, Hong KH, Werner MD, Upham KM, Treff NR, et al. Aneuploidy across individual chromosomes at the embryonic level in trophoctoderm biopsies: changes with patient age and chromosome structure. *J Assist Reprod Genet* 2014;31:1501–9.
 212. Bajpai K, Acharya N, Prasad R, Wanjari MB. Endometrial receptivity during the preimplantation period: a narrative review. *Cureus* 2023;15:e37753.
 213. Winstanley YE, Stables JS, Gonzalez MB, Umehara T, Norman RJ, Robker RL. Emerging therapeutic strategies to mitigate female and male reproductive aging. *Nat Aging* 2024;4:1682–96.
 214. Kumar M, Kumar K, Jain S, Hassan T, Dada R. Novel insights into the genetic and epigenetic paternal contribution to the human embryo. *Clinics (Sao Paulo)* 2013;68:5–14.
 215. Practice Committees of the American Society for Reproductive Medicine and the Society for Assisted Reproductive Technology. The use of preimplantation genetic testing for aneuploidy: a committee opinion. *Fertil Steril* 2024;122:421–34.
 216. Practice Committee of the American Society for Reproductive Medicine. Practice Committee of the Society for Assisted Reproductive Technology. Blastocyst culture and transfer in clinically assisted reproduction: a committee opinion. *Fertil Steril* 2018;110:1246–52.